

HAEMATOLOGY

ANALYSIS OF THE BLOOD CELLS AND INTERPRETATION OF RESULTS

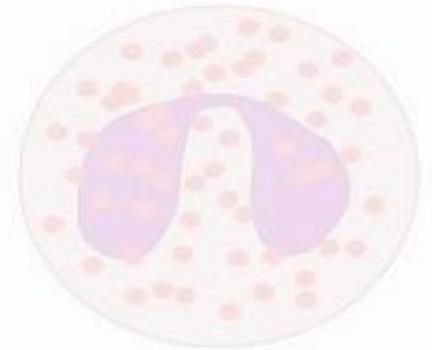
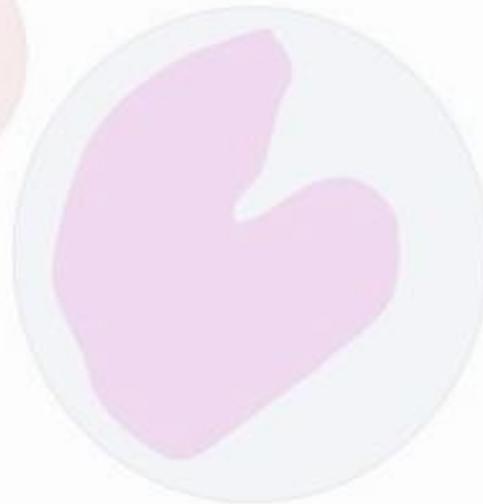
Gabriella Kiss

19th september 2019

2nd practice

PARAMETERS OF CBC

- **WBC** (white blood cell count, Giga/l) **5-10 G/l**
 - **NEU%**, **NEU#** (neutrophil granulocyte) **40-70%**
 - **LY%**, **LY#** (lymphocyte) **20-40%**
 - **MO%**, **MO#** (monocyte) **4-8%**
 - **EO%**, **EO#** (eosinophil granulocyte) **2-4%**
 - **BAS%**, **BAS#** (basophil granulocyte) **0,5-1%**



Relative lymphocytosis

Vérkép automatával:

Fehérvérsejt	2,500	L	Giga/l	4,000-10,000
<u>Neutrofil</u>	<u>27,4</u>	L	%	34,0-71,1
<u>Neutrofil (abs)</u>	<u>0,69</u>	L	Giga/l	1,56-6,13
<u>Limfocita</u>	<u>65,9</u>	H	%	19,3-51,7
<u>Limfocita (abs)</u>	<u>1,65</u>		Giga/l	1,18-3,74
Monocita	4,9	U	%	4,7-12,5
Monocita (abs)	0,120	U~L	Giga/l	0,240-0,860
Eozinofil	1,3	U	%	0,0-5,8
Eozinofil (abs)	0,030		Giga/l	0,000-0,360
Bazofil	0,6		%	0,0-1,2
Bazofil (abs)	0,010		Giga/l	0,000-0,080
Vörösvértest	2,66	U~L	T/l	3,90-5,30
Hemoglobin	86	U~L	g/l	120-157
Hematokrit	24,1	U~L	%	34,1-44,9
MCV	90,6		fl	80,0-95,0
MCH	32,3		pg	26,0-33,0
MCHC	356		g/l	310-360
RDW	16,1	H	%CV	11,6-14,4
Trombocita	36,0	L	Giga/l	140,0-440,0
MPV	6,55	L	fl	9,40-12,40

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 - **EO%**, **EO#** (eosinophil granulocyte) **2-4%**
 - **BAS%**, **BAS#** (basophil granulocyte) **0,5-1%**
- **RBC** (red blood cell count, Tera/l) **4,1-5,1/4,5-5,9 T/l**
- **HGB** (haemoglobin, g/l) **123-153/140-175 g/l**
- **HCT** (haematocrit, %) **35-43/38-49 %**
- **MCV** **80-95 fl**
- **MCH** **26-33 pg**
- **MCHC** **310-360 g/l**
- **RDW-CV** **11,6-14,4%**
- **PLT** (platelet, Giga/l) **150-400 G/l**
- **MPV** (mean platelet volume)-fl

CALCULATED PARAMETERS

MCV= HTC/RBC (mean volume of red blood cells in fl)*

MCH= HGB/RBC (mean corpuscular hemoglobin in pg)

MCHC= HGB/HTC (mean corpuscular hemoglobin concentration in g/l)-the hemoglobin concentration of 1 l red cell mass

RDW -CV% (11,6-14,4)

RDW-SD (fl)

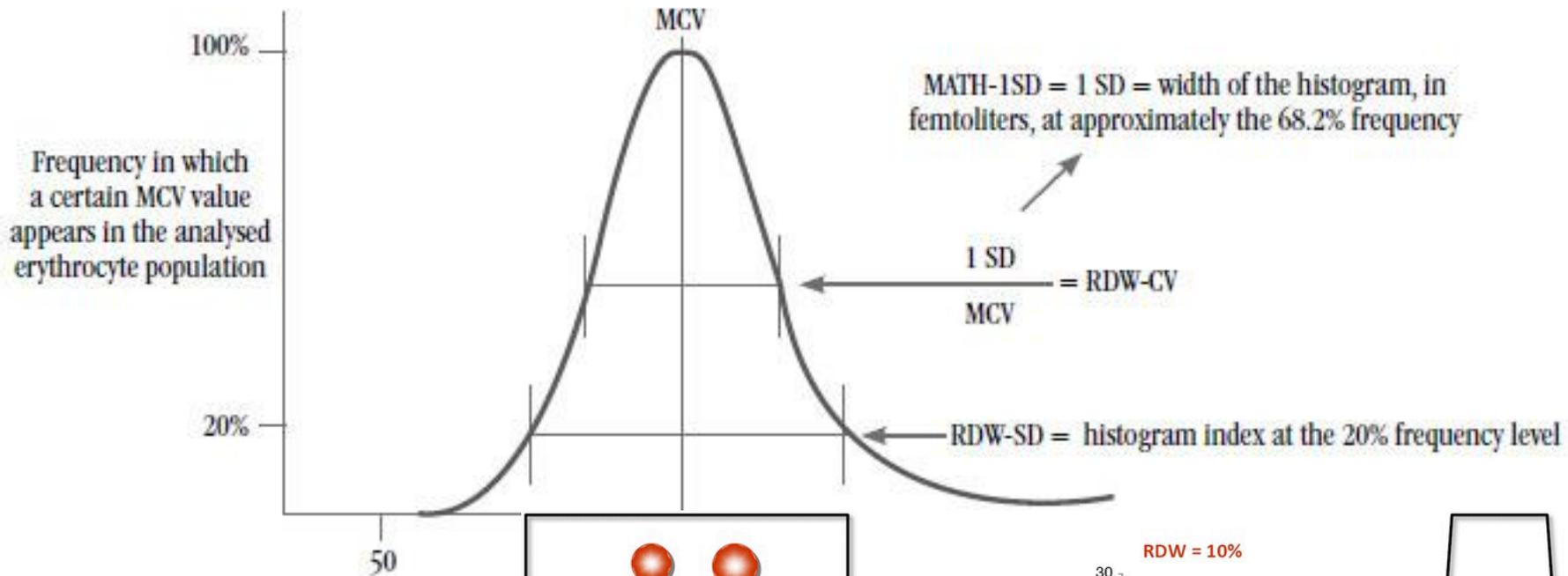
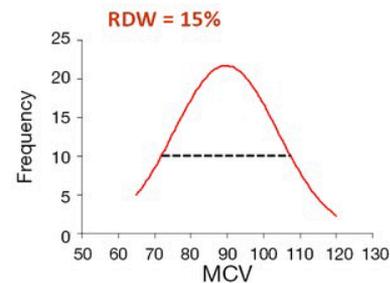
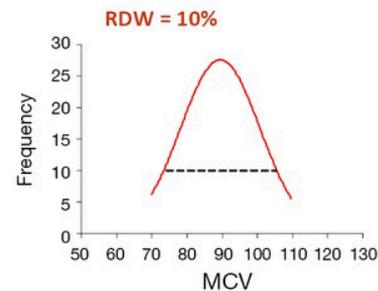
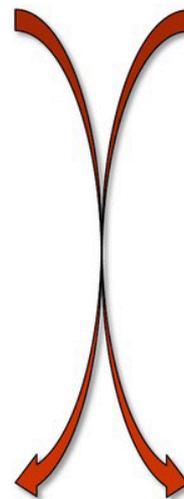
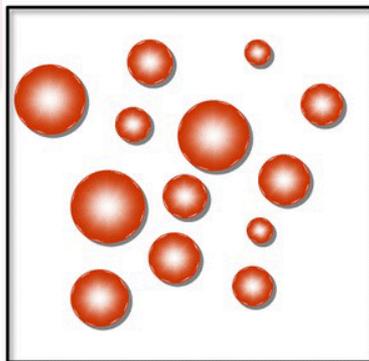
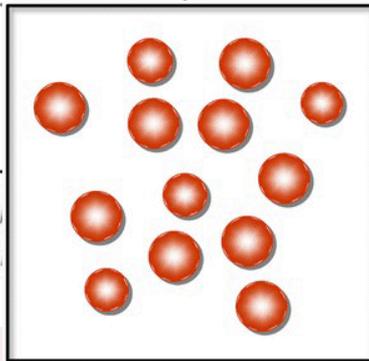


FIGURE – *Obtainment of RDW-CV, MATH-1SD and RDW-SD*
RDW-CV: coefficient of variation of red cell distribution width; volume.

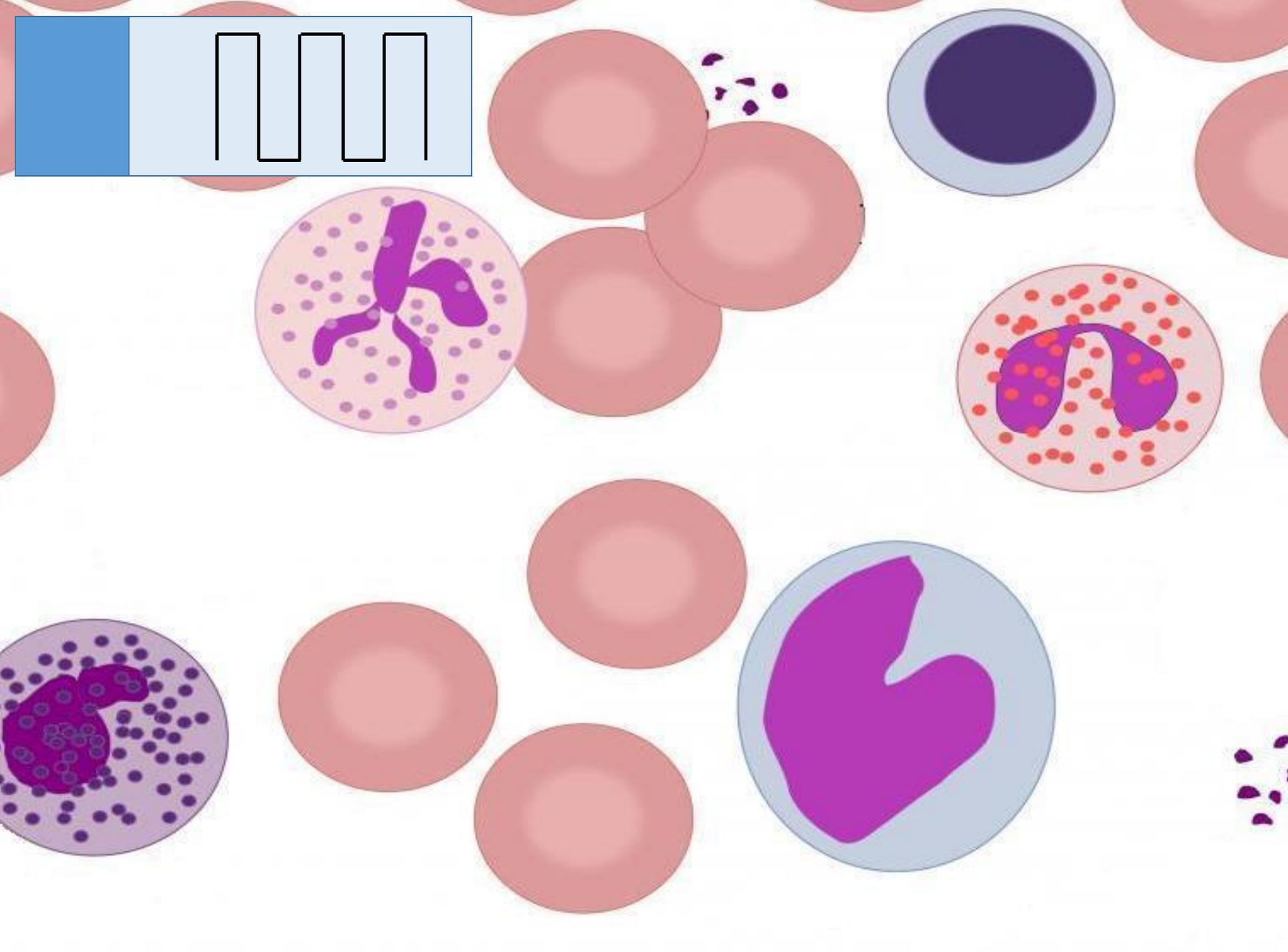
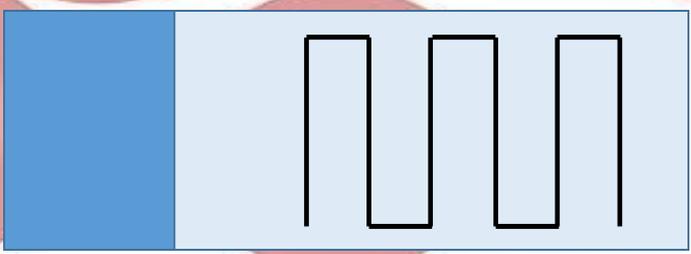


Anisocytosis

PREANALYTICS

- ▣ K3-EDTA, lavender tube, mixing well
- ▣ posture (getting up: HGB, HCT increase by 10% or more)
- ▣ must be measured within 4 hours, but as fast as possible except for: reticulocyte (within 24 hrs in room temp., 72 hrs if stored at 4°C)
- ▣ peripheral smear procedure:
 - capillary blood from fingertip, or from EDTA-tubes within 2-3 hours





BLOOD CELLS

white blood cells:

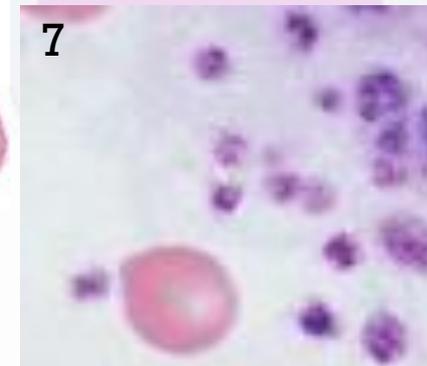
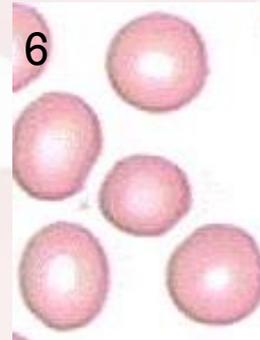
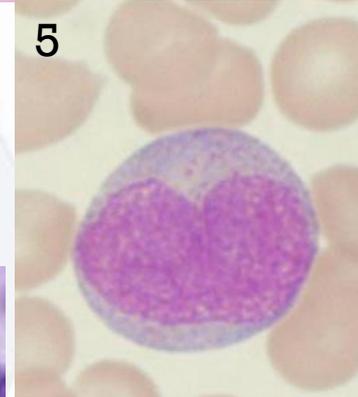
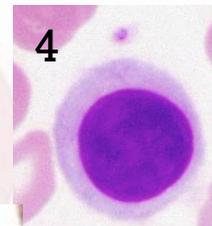
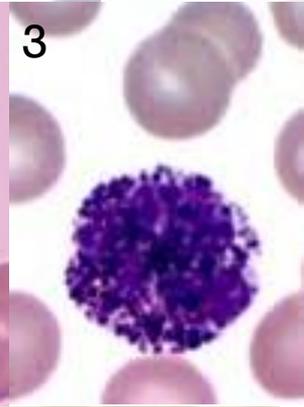
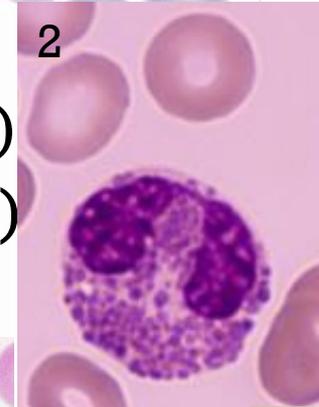
1. neutrophil granulocyte (10-12 μm)
2. eosinophil granulocyte (10-12 μm)
3. basophil granulocyte (10-12 μm)
4. lymphocyte (9 μm)
5. monocyte (15-30 μm)

red blood cells:

- diameter: 7-8.5 μm
- thickness: 2 μm

platelets:

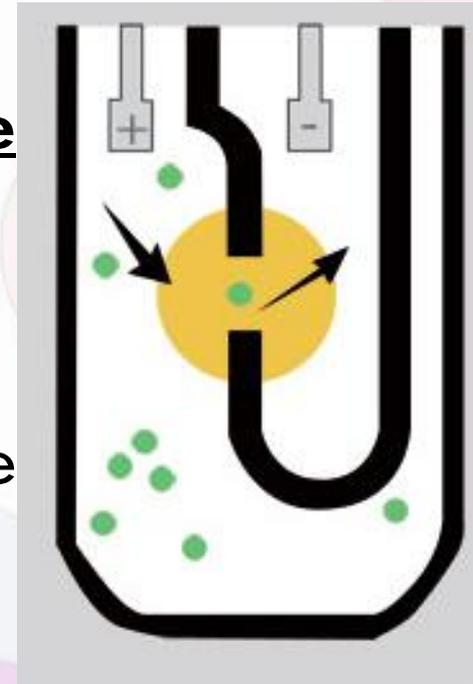
- diameter: 2-5 μm



HAEMATOLOGICAL INSTRUMENTS: PRINCIPLES 1.

Coulter-principle (Wallace Coulter-1953)

- detection of changes in electric impedance
- orifice, between its sides DC (direct electric current)–gradient is present
- the sample is diluted with a good conductor liquid, when a cell passes through the orifice electrical resistance increases → current intensity decreases ($U=IxR$)
- the size of the current change is related to the size of the cell, the number of it related to the number of the cells (volume-distribution histogram)

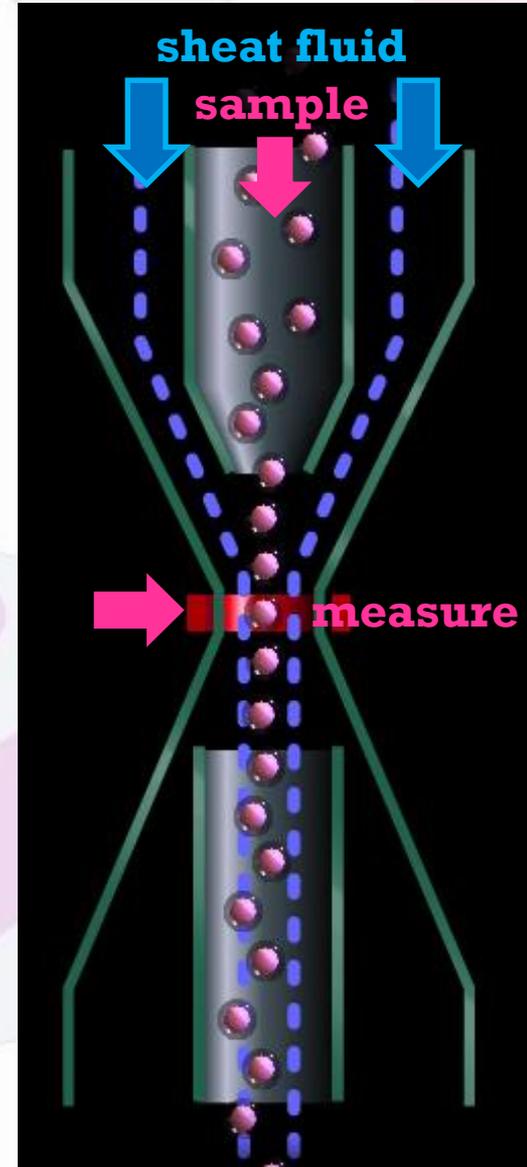


Sources of error:
• coincidence
• recirculation

HAEMATOLOGICAL INSTRUMENTS: PRINCIPLES 2.

hydrodynamic focusing

- to eliminate coincidence and recirculation
- the created laminar flow (sheat fluid) permits an unilinear pass of the cells through the site of measurement
- the constant flow decreases the chance of clog formation (proteins)



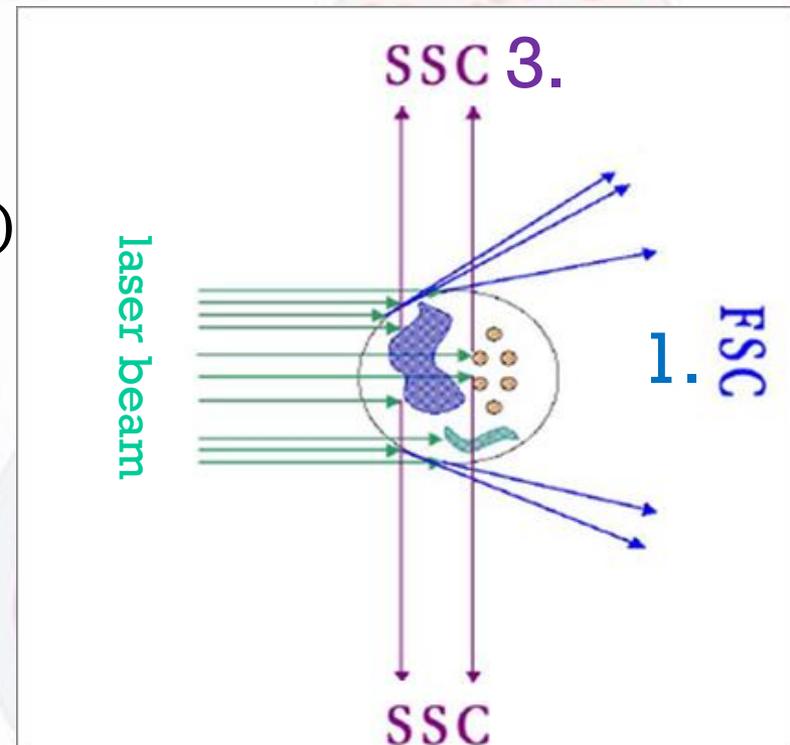
HAEMATOLOGICAL INSTRUMENTS: PRINCIPLES 3.

optical principal

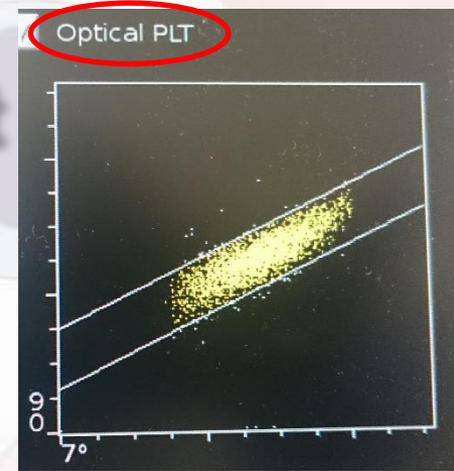
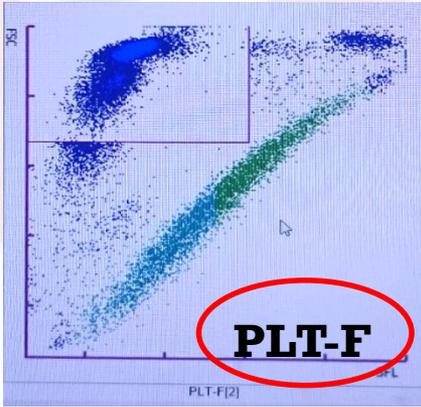
- the refractive index of cells differs from the diluent solution
- argon/helium laser beam
 1. forward scatter detector ($1-3^\circ$): size/volume of cells (scattering light)
 2. low angle scatter detector ($7-11^\circ$): indicator of cells' complexity
 3. side scatter detector ($70-110^\circ$): lobularity of the cells (refraction and reflexion of the light)

other specific prospects

- special stains (fluorescent, cytochemical)
- special characteristics of cells (selective lysis, depolarization, etc.)



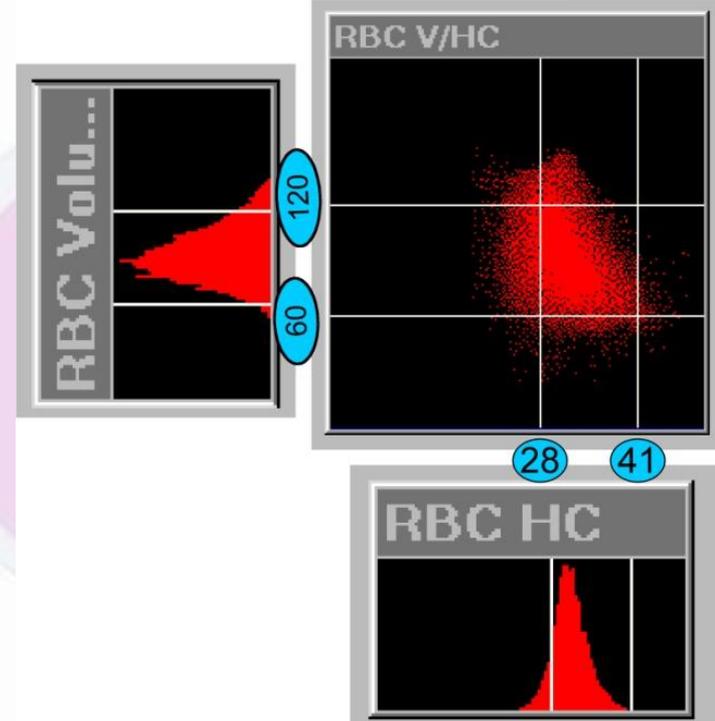
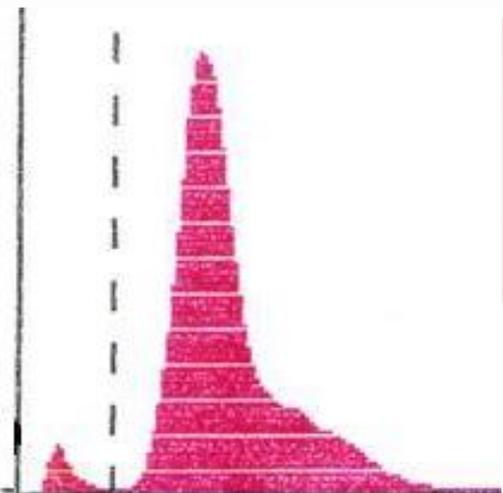
Red cell and platelet measurements



- ▣ Coulter-principle (impedance) supplemented with optical method
- ▣ FSC: volume (MCV)
- ▣ SSC: haemoglobin content

RBC volume/haemoglobin content histogram and dot-plot based on light scatter

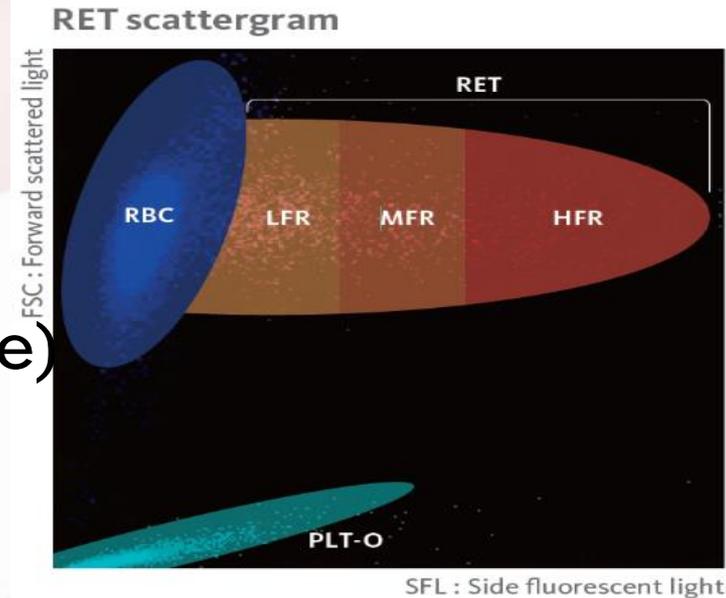
WBC-do we have to remove them for measuring RBC-s?



PLT-RBC volume distribution histogram

COUNTING RETICULOCYTES

- ▣ last stage of premature RBCs (no nucleus, contains RNP)
- ▣ 1-1,5 days in periphery before becoming mature RBC
- ▣ using supravital stain (brilliant cresyl blue, methylene-blue)
- ▣ analyser:
 - nucleic acid stain (fluorescent)
 - IRF (immature reticulocyte fraction)
 - CHr (haemoglobin content of reticulocyte)-iron deficiency
- ▣ reticulocyte crisis (anaemia of vitamin B12, folic acid deficiency 24 hours after treatment)-IRF increases
- ▣ information about bone marrow function

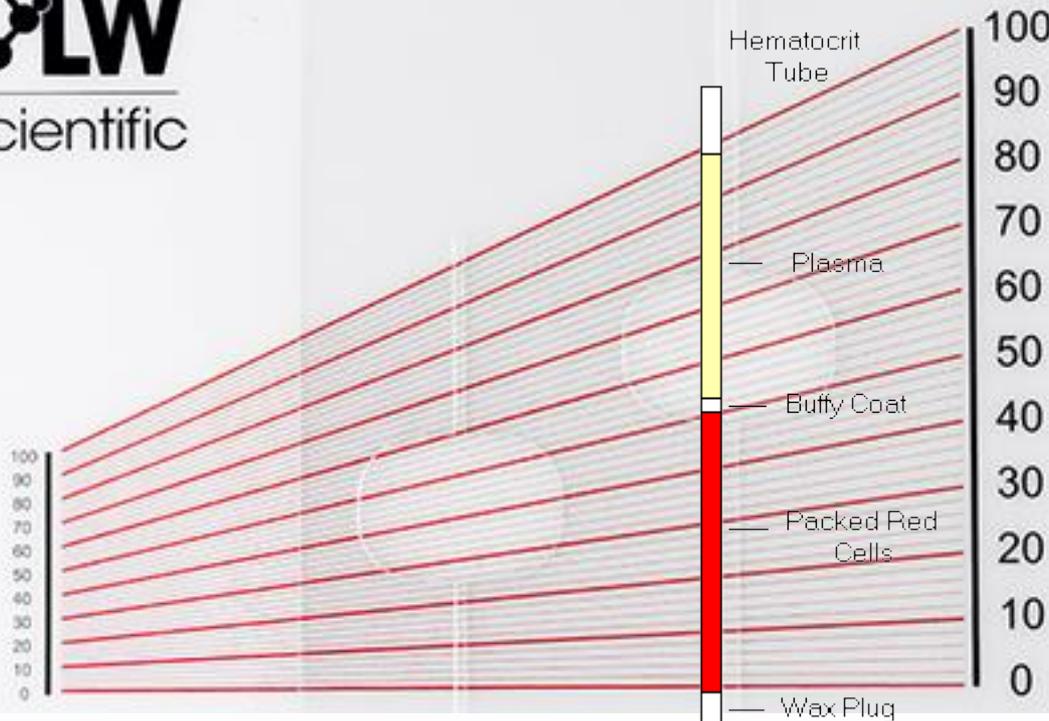


HAEMATOCRIT MEASUREMENT

▣ the past



LW
Scientific



EZ Reader

Microhematocrit Reader
for 40mm and 75mm capillary tubes

TO USE:

1. Place centrifuged tube in slot with interface of sealant and packed red cells intersecting the '0%' line.
2. Slide the tube holder until the top of plasma layer intersects the '100%' line.
3. Read the percentage height of the red cell column from the scale.

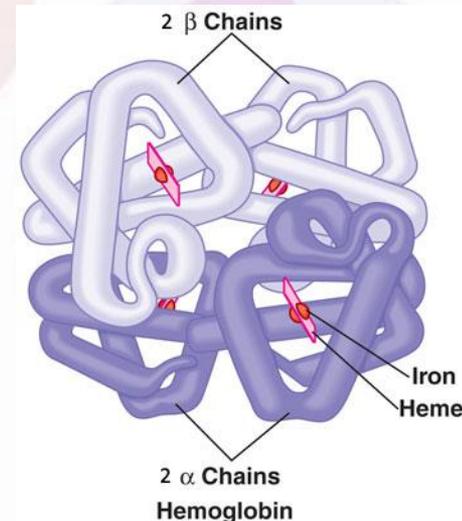
Lines are in 2% increments, therefore estimate to the nearest 1%.

For *in vitro* diagnostic use.



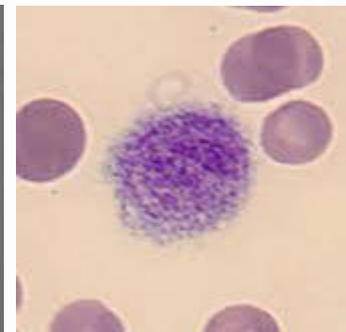
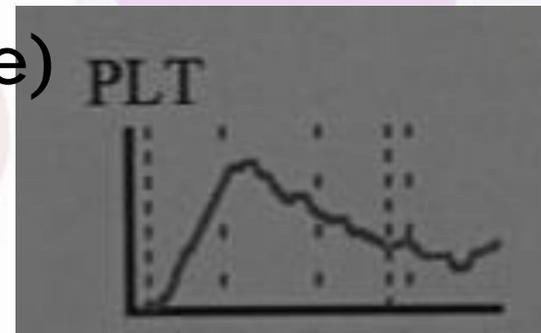
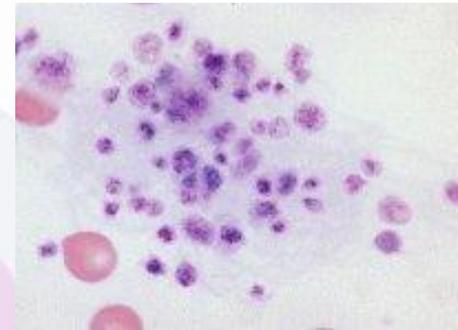
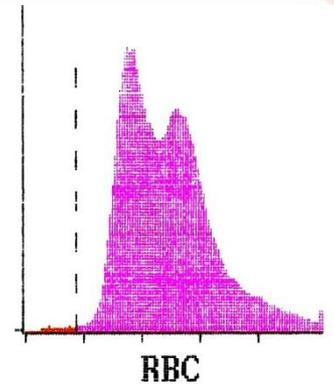
HAEMATOCRIT, HAEMOGLOBIN MEASUREMENTS

- ▣ haematocrit: the present
 - analyser: $PCV = MCV \times RBC$
- ▣ haemoglobin:
 - photometric detection
 - stable hemoglobin complex (SLS, (cyanid))
 - interfering factors:
 - lipaemic samples
 - increased bilirubin
 - (cold-agglutinins)
 - (hemolysis)



Transfusion, pseudothrombocytopenia, plt aggregates, cold agglutinin

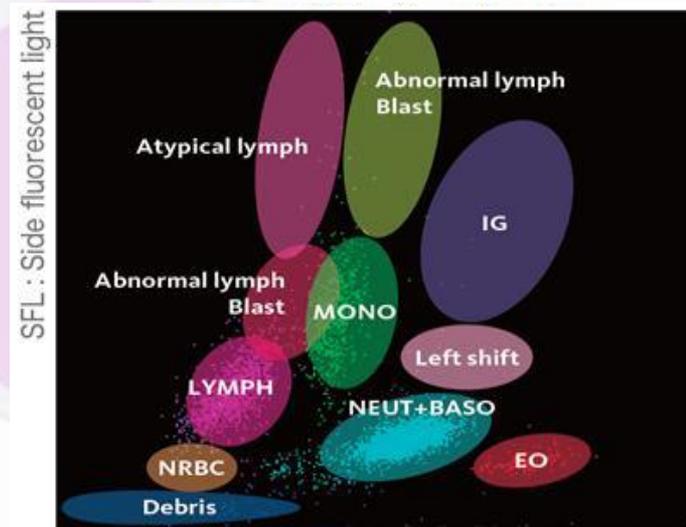
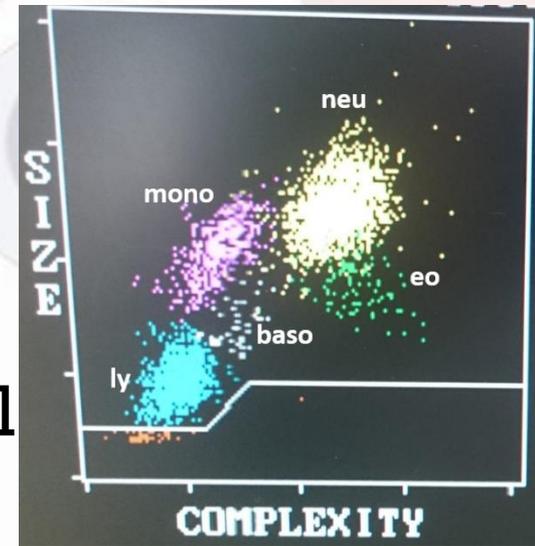
- transfusion/recovery from pernicious anaemia, or iron deficiency anaemia: two populations of rbc-s
- pseudotrombocytopenia: when plt count~0 from EDTA tube, normal from citrate tube, blood smear: tct aggregates
- giant platelets (impedance)
- cold agglutinin:
repeat measurement at 37°C



RBC-do we have to remove them for measuring WBC-s?

WBC COUNT

- ▣ site of measurement: optical flow cell
- ▣ dot-plots
- ▣ 5 part diff
- ▣ 6 part diff:
 - IG (immature granulocytes)



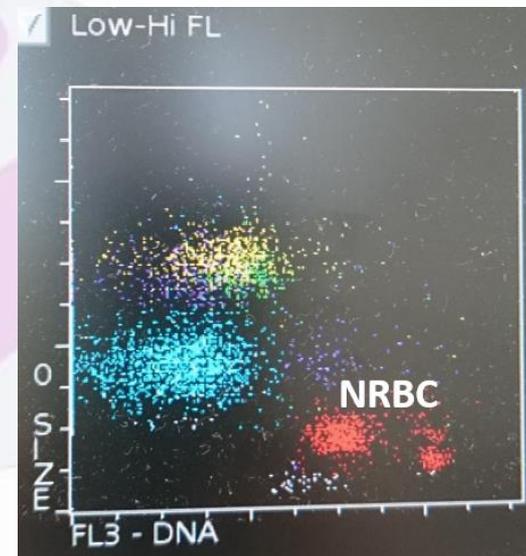
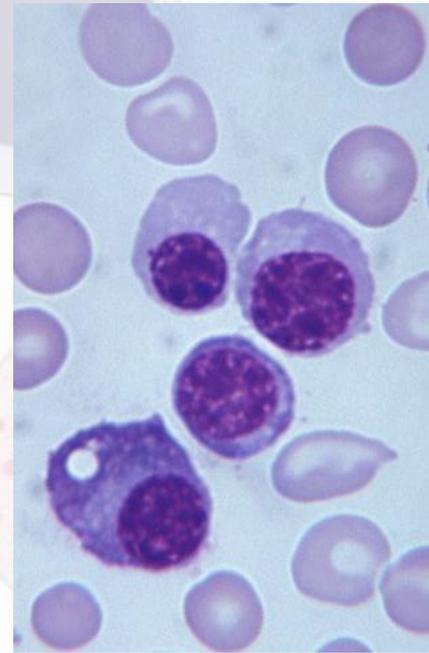
SSC : Side scattered light

FLAGS (notifications)

- ▣ suspicious population of cells because of inadequate separation (DIFF)
- ▣ cells at abnormal site at the dot-plot (IG/BAND, BLAST/VARLYMPH)
- ▣ plt aggregates (PLTR), nucleic rbc-s (NRBC)
- ▣ smear should be examined:
suspicion of presence of malignant cells,
or measuring error
- ▣ nucleic RBC-s

Nucleic red blood cells

- ▣ normally they are in the bone marrow
- ▣ common in newborn
- ▣ in some disease it can be found in adults
- ▣ can falsely determined as lymphocytes (high WBC, high ly % and abs. count)
- ▣ methods to count them:
 - nucleic acid stain (e.g. propidium-iodide)



ANAEMIAS

- ▣ **definition: reduced red cell mass**
- ▣ **according to red cell morphology or MCV, MCH:**
 - normochrom normocytic
 - hypochrom microcytic
 - hyperchrom macrocytic
- ▣ **based on etiology:**
 - decreased production (bone marrow)
 - deficiency anemias
 - shortened lifespan (hemolysis, inherited diseases, kidney failure...)
 - increased loss (occult bleeding, acute bleeding)
 - dilution (pregnancy)

Vérkép automatával:

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Monocita (abs)	0,330		Giga/l	0,240-0,860
Eosinofil	1,0		%	0,0-5,8
Eosinofil (abs)	0,060		Giga/l	0,000-0,360
Basofil	0,4		%	0,0-1,2
Basofil (abs)	0,020		Giga/l	0,000-0,080
Gép által nem azonosítható FVS	1,700		%	<5,000
RBC	5,77	H	T/l	3,90-5,30
HGB	124		g/l	120-157
Haematocrit	40,0		%	34,1-44,9
MCV	69,3	L	fl	80,0-95,0
MCH	21,5	L	pg	26,0-33,0
MCHC	310		g/l	310-360
RDW	20,6	H	%CV	11,6-14,4
Trombocita	206,0		Giga/l	140,0-440,0
MPV	6,90	L	fl	9,40-12,40

DIAGNOSING ANAEMIAS

- ▣ **CBC** (RBC, HGB, HTC, MCV, MCH, MCHC, RDW)
- ▣ **reticulocyte**
- ▣ **iron parameters** (iron, ferritin, transferrin, transferrin saturation, soluble transferrin receptor, bone marrow iron storage-Prussian-blue)
- ▣ **CRP**
- ▣ **blood smear**
- ▣ **deficiencies:** serum vit. B12, folic acid
- ▣ **in the case of haemolysis:** LDH, indirect bilirubin, urine UBG, haptoglobine
- ▣ **special investigations:** haemoglobin elpho, genetical investigation, Coombs-test

THROMBOCYTE DISORDERS

▣ thrombocytopenia:

- under 50 Giga/l risk for bleeding
- decreased production: inherited (Fanconi anaemia, May-Hegglin anomaly)/acquired (drugs, tumours, hematological malignancies, radiotherapy)
- increased destruction: ITP, TTP, HUS
- hypersplenism (increased storage)

▣ thrombocytosis:

- ET, MDS
- after splenectomy transiently, chronic inflammation, tumor, iron deficiency, hemolytic anaemia

▣ qualitative disorders:

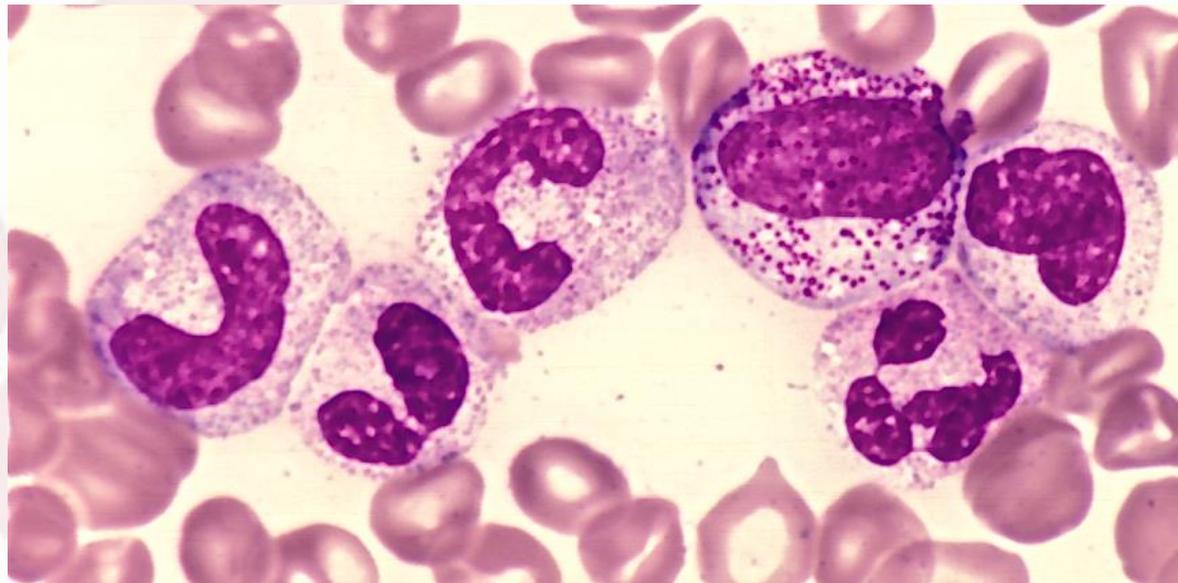
- acquired (ASA, uraemia, liver disease)/inherited (Bernard-Soulier syndrome, Gray platelet syndrome)

Ultraszenzitiv CRP	18,72	H	mg/l	<5,00
Ultraszenzitiv CRP	8,27	H	mg/l	<5,00
Ultraszenzitiv CRP #	4,99		mg/l	<5,00
Vérkép automatával:				
Fehérvérsejt	3,530	L	Giga/l	4,000-10,000
Neutrofil	35,2		%	34,0-67,9
Neutrofil (abs)	1,24	L	Giga/l	1,78-5,38
Limfocita	46,9		%	21,8-53,1
Limfocita (abs)	1,65		Giga/l	1,32-3,57
Monocita	13,6	H	%	5,3-12,2
Monocita (abs)	0,479		Giga/l	0,300-0,820
Eozinofil	3,9		%	0,0-7,0
Eozinofil (abs)	0,138		Giga/l	0,000-0,540
Basofil	0,4		%	0,0-1,2
Basofil (abs)	0,016		Giga/l	0,000-0,080
Vörösvértest	4,84		T/l	4,50-6,00
Hemoglobin	140		g/l	137-175
Hematokrit	41,2		%	40,1-51,0
MCV	85,1		fl	80,0-95,0
MCH	28,9		pg	26,0-33,0
MCHC	340		g/l	310-360
RDW	16,2	H	%CV	11,6-14,4
Trombocita	127,0	U~L	Giga/l	140,0-440,0
MPV	6,96	L	fl	9,40-12,40
Retikulocita HG tartalom #	25,3	L	pg	28,0-35,0

NEUTROPHILIA, LEUKAEMOID REACTION

- ▣ neutrophilia: stress, adrenaline, corticosteroids, pregnancy, infection, inflammation
- ▣ leukaemoid reaction: present in inflammation, important to differentiate from CML (granulocyte alkaline phosphatase=GAP)

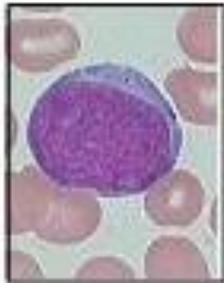
toxic granules



left shift, younger stages of maturation appear in periphery

Maturation of granulocytes

myeloblast promyelocyte myelocyte metamyelocyte band neutrophil



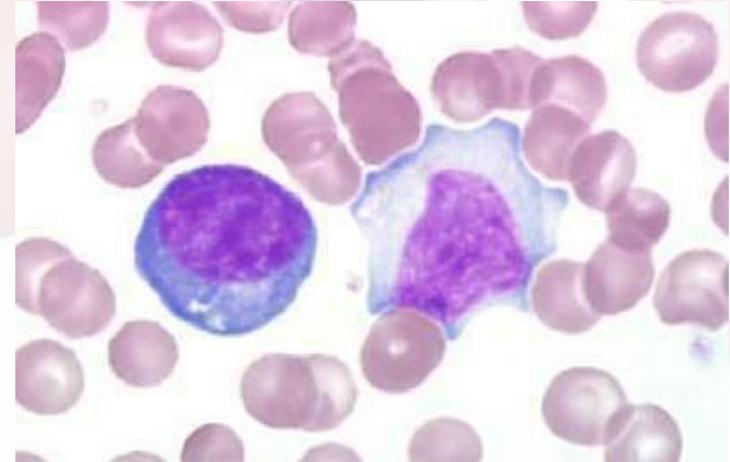
neutrophilic-
eosinophil-
basophilic-
myelocyte

division

maturation

MONONUCLEOSIS INFECTION

- ▣ caused by EBV
- ▣ fever
- ▣ adenomegaly (usually in the neck),
- ▣ hepatosplenomegaly
- ▣ elevation of liver enzymes, because of hepatitis
- ▣ presence of atypical mononuclear cells (usually lymphocytosis, maybe monocytosis, when measured with the analyser)

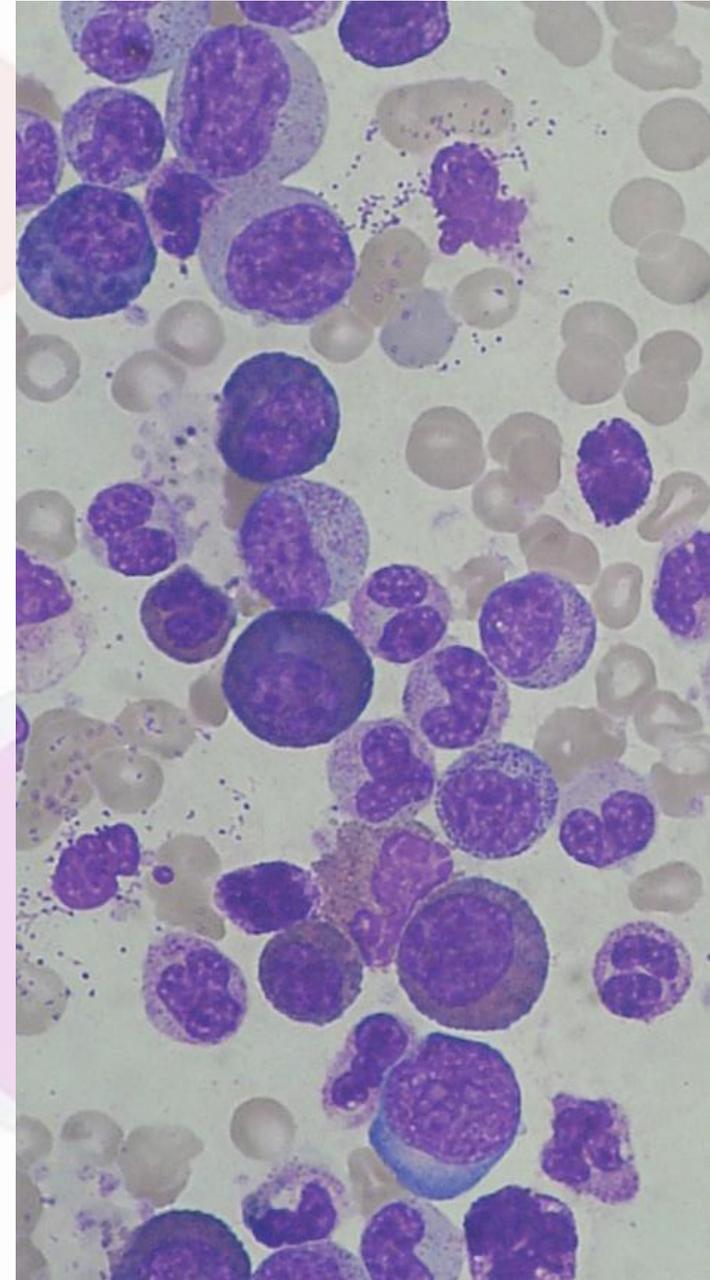


LEUKAEMIAS

- ▣ =higher numbers of abnormal WBCs
- ▣ blast cells (stem cells present normally in the bone marrow, less than 5%)
- ▣ the clonal expansion of the cells results from mutations in the mother cell's DNA
- ▣ depending on origin it can be lymphoid or myeloid
- ▣ depending on the number of blasts it can be acute (more than 20%) or chronic

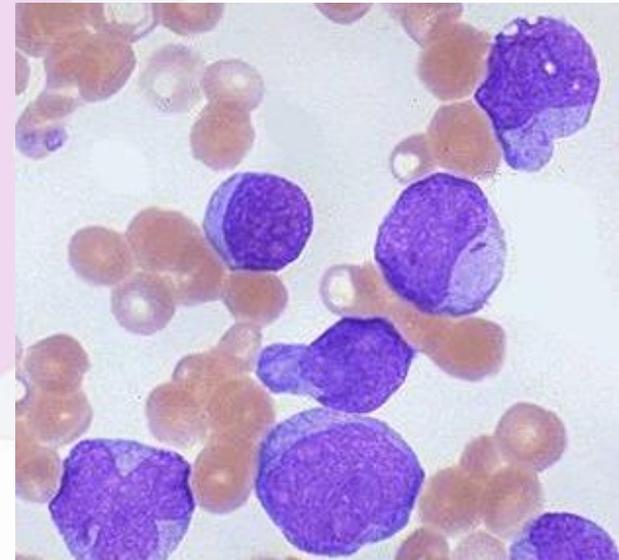
CHRONIC MYELOID LEUKAEMIA (CML)

- ▣ high WBC
(more than 50 Giga/l)
- ▣ in the periphery all the maturation stages present
- ▣ decreased GAPA activity, Philadelphia chromosome (BCR-ABL fusion gene)
- ▣ can transform to acute leukaemia



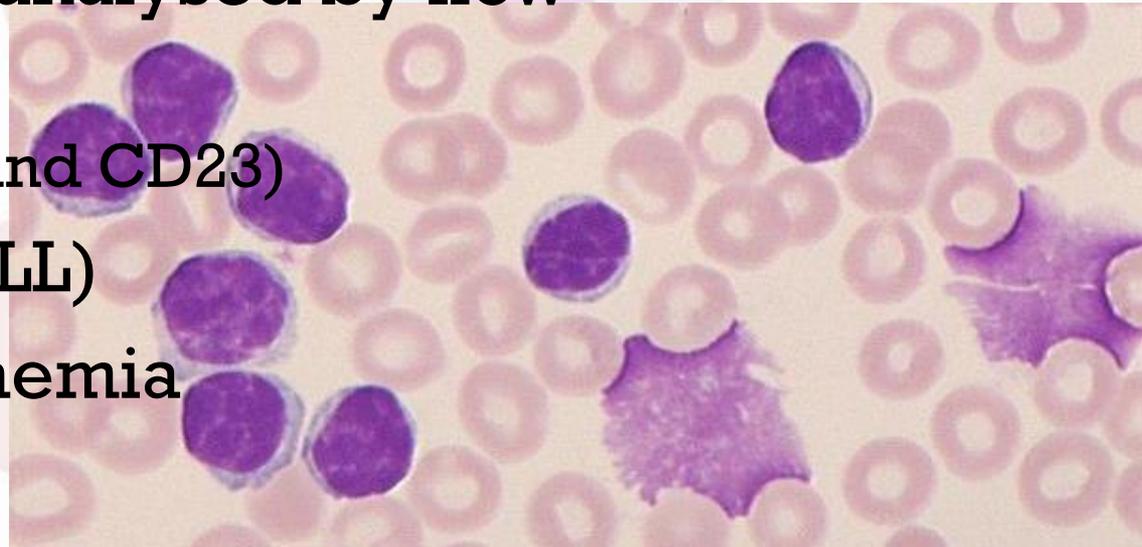
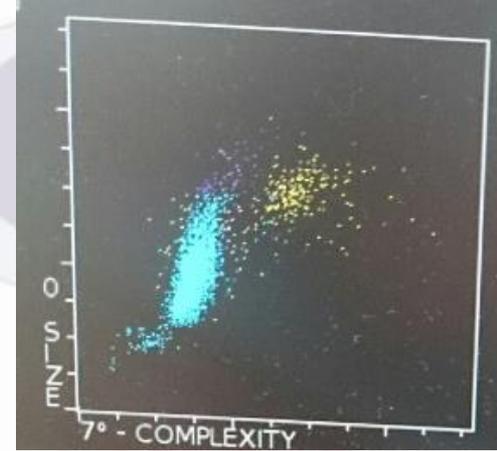
ACUTE MYELOID LEUKAEMIA (AML)

- ▣ in the bone marrow or in the periphery the rate of blast cells increase up to more than 20%
- ▣ hiatus leukaemicus
- ▣ in the blasts Auer-rods can be present
- ▣ usually positive with MPO stains (except for the completely immature blast cells)
- ▣ exact diagnosis: flow cytometry



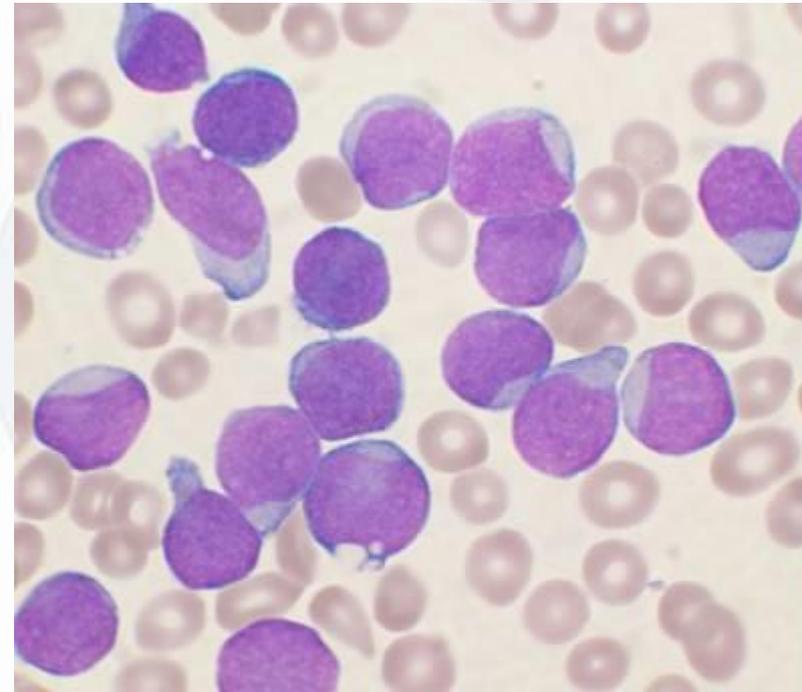
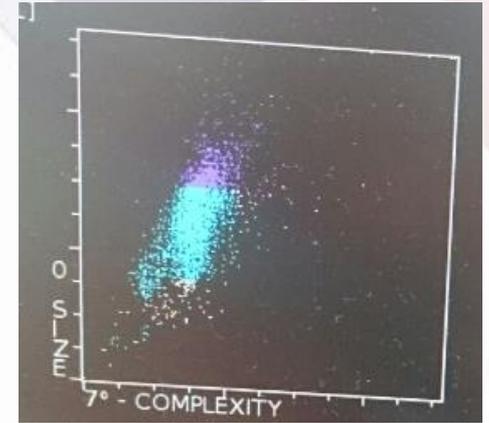
CHRONIC LYMPHOID LEUKAEMIA (CLL)

- ▣ the appearance of small mature lymphocytes in the peripheral blood (having heterochromatic nucleus)
- ▣ cells are fragile, when spreading the smear
- ▣ typical phenotype analysed by flow cytometry
(positive with both CD5 and CD23)
- ▣ can transform to (PLL)
prolymphocytic leukaemia



ACUTE LYMPHOID LEUKAEMIA (ALL)

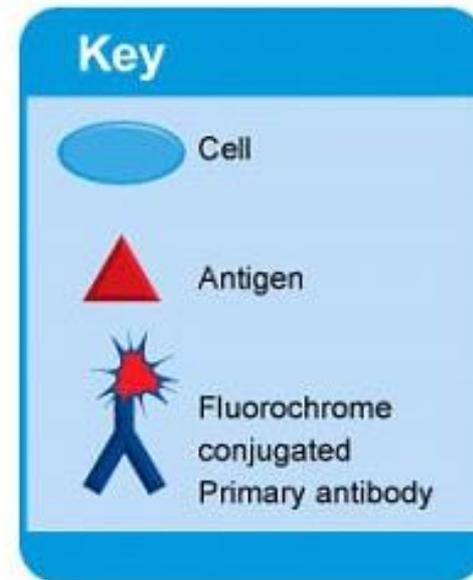
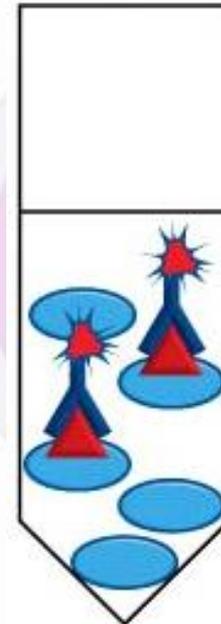
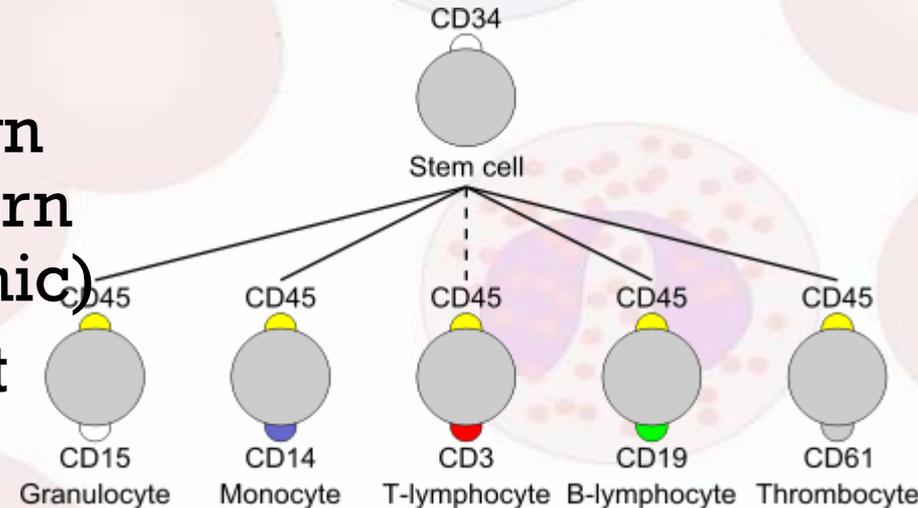
- ▣ increase in number of lymphoblastic cells (from bone marrow or extramedullar organs)
- ▣ most common type of cancer in children
- ▣ classified based on immunophenotyping and genotyping (low risk/high risk)



Flow cytometry 1.

BASICS:

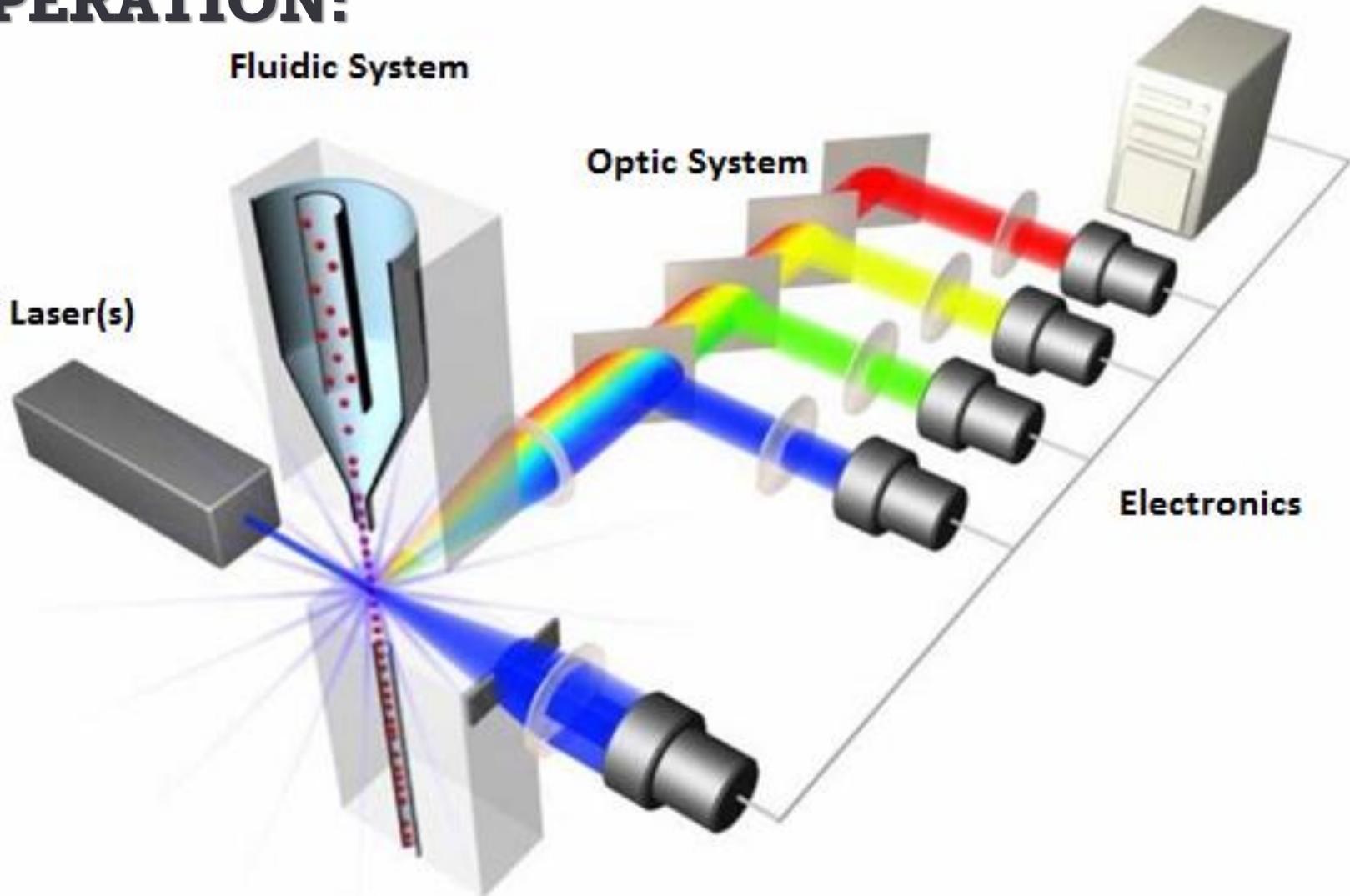
- every type of cell has its own individual CD-marker pattern (surface and intracytoplasmic)
- adding the sample different monoclonal antibodies conjugated with different coloured fluorescent dyes result in specific binding of antibodies by the cells
- then the fluorophores are excited by a laser beam, after it the intensity of emission will be measured



Flow cytometry 2.

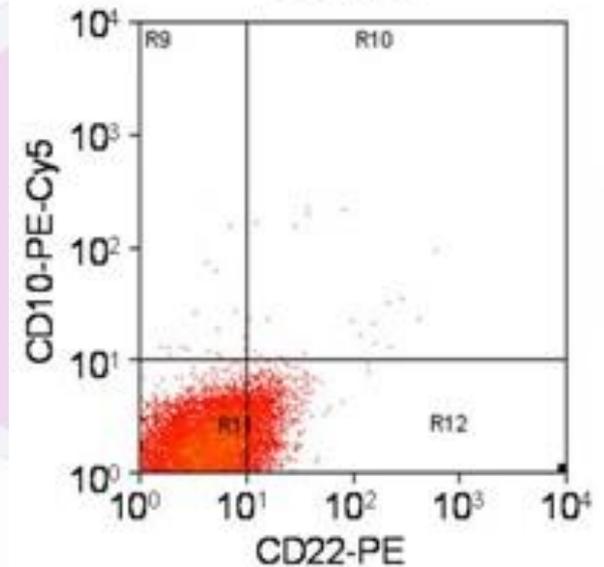
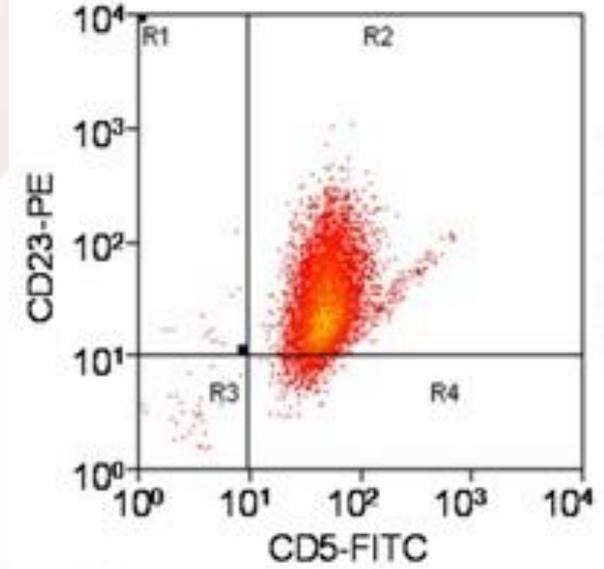
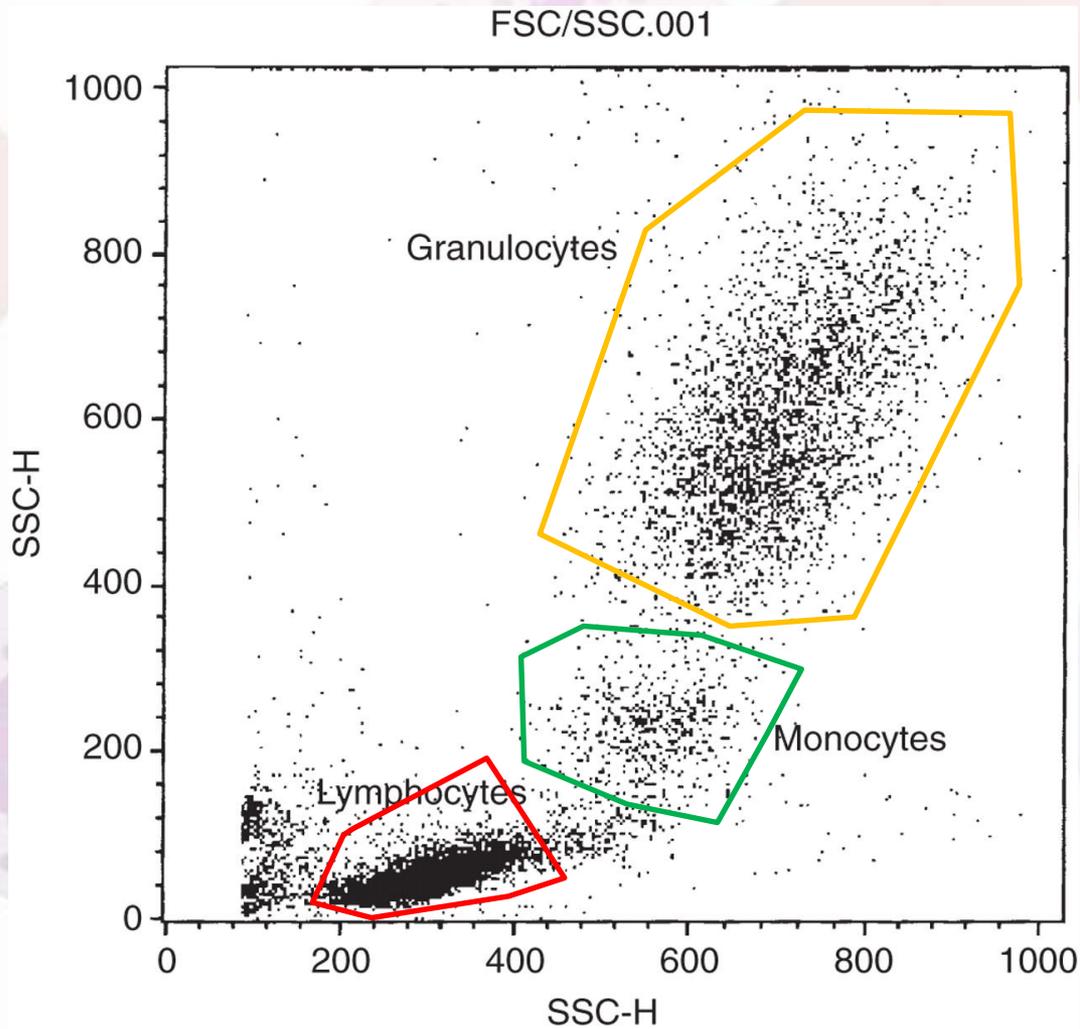
OPERATION:

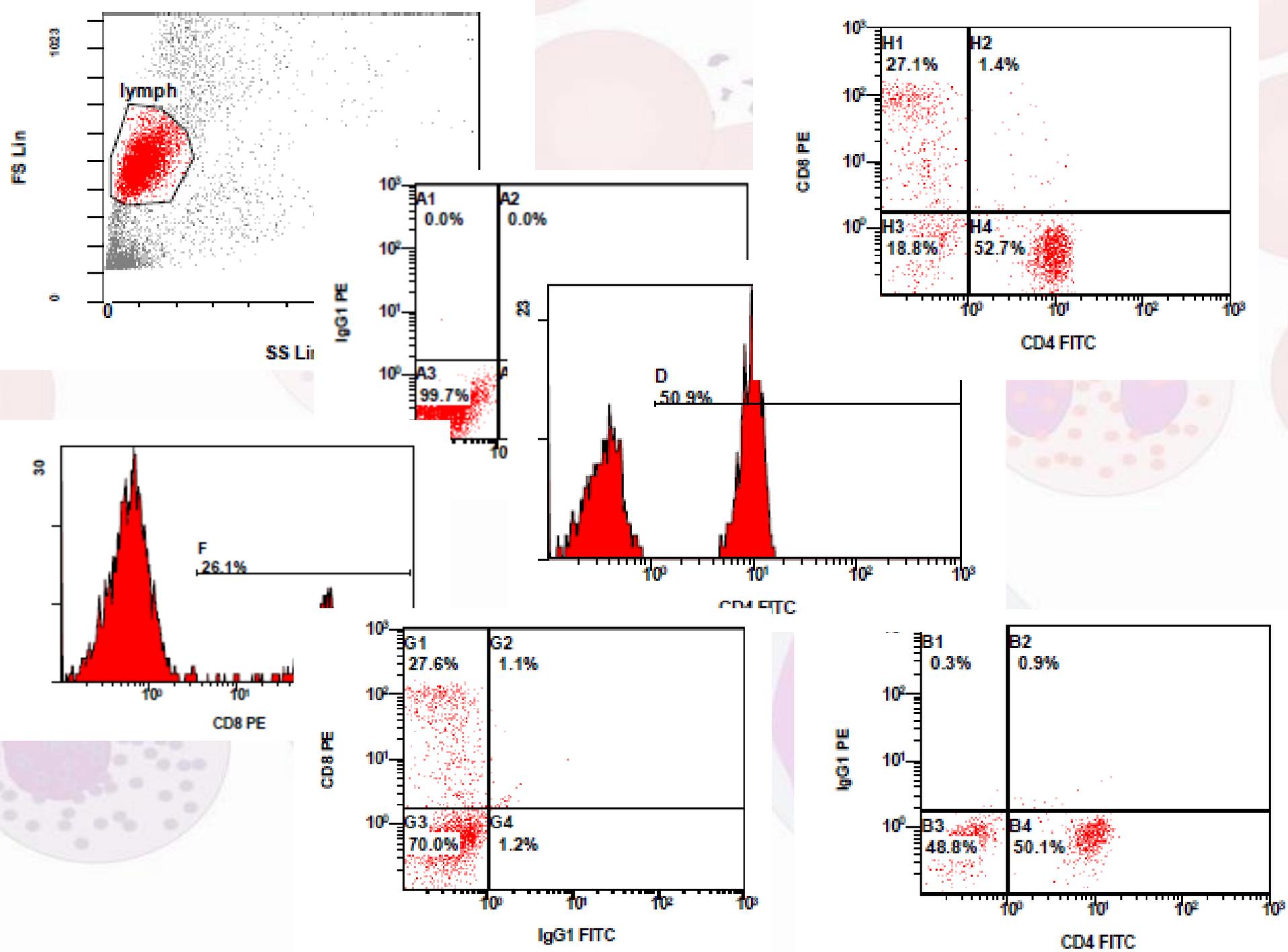
Fluidic System



Flow cytometry 3.

EVALUATION:





CASE STUDY

- ▣ 5 y.o. boy
- ▣ somnolent, exhaustible for a week, subfebrility
- ▣ started to vomit and had fever from a few days (38,2 °C)
- ▣ from that he cannot be fed, vomited again
- ▣ stool and urine was ok
- ▣ practitioner: hepatosplenomegaly
- ▣ requesting blood test

Vizsgált szerv:

perifériás vér

Minta típusa:

Natív Anyag, Kenetek

Klinikai diagnózisok:

ALL in obs

Klinikai adatok:

Háziorvosánál 02.27-én jelentkezett subfebrilitás és fáradékonyság miatt, hepatosplenomegáliát véleményezett és laboratóriumi vizsgálatot kért, mely eredményének tükrében (fvs: 1,6 millió, HgB: 66, Htk: 19, tct: 36000) kéri osztályos felvételét.

Panaszai 1 hete szerdán kezdődtek (2012.02.21-én). Fáradékonyságot, alusszékonyságot észleltek nála. Hőemelkedése jelentkezett. Pénteki napon hányt először. Hétvégén panaszai fokozódtak, vasárnap beléhasodott (38,2°C). Hányás ismét jelentkezett, mai napon nem volt per os táplálható, az elfogyasztott fél liter folyadékot is kihányta. Székletet szombaton ürített utoljára, kis mennyiségűt, normál állagút. Mai napon 1 alkalommal ürített kevés vizet, mely nem véres, nem barna, nem csíp.

Makroszkópos leírás

A: 3ml EDTAs vér, B: 5db perif kenet (saját), C: 13db perif kenet (küldött)

Mikroszkópos leírás

02.29.

A kenetekben extrém leukocytosis figyelhető meg, a sejtek 98%-a lymphoblast, melyek többsége közepes méretű, a sejtek peroxidás negatív cytoplasmával rendelkeznek.

A minta áramlási citometriai vizsgálata során 4% myeloid és 2% lymphoid sejt mellett 94% blast karakterű sejt mutatható. A blastok CD58, CD38, CD99, CD19 expressziót mutattak, 30%-ban CD34 jelölődés, 6%-ban minimális intenzitású cytoplasmicus IgM expresszió mellett. A sejtek CD20, CD10 és TdT negatívak, a sejtfelületen könnyűláncok nem mutathatók, myeloid, illetve T-sejtes markerek nem voltak jelen.

Diagnózisok:

ACUT LYMPHOBLASTOS LEUKAEMIA

CD19+, CD38+, CD58+, CD99+, CD20-, TdT-, CD10-, CD34-/+

BNO: C9100 Heveny lymphoblastos leukaemia

**THANK YOU FOR YOUR
ATTENTION!**

